

COVID-19+Flu A&B+RSV Antigen **Combo Rapid Test Cassette** (Nasopharyngeal swab) Package Insert REF IRCF-MC84 English

A rapid test for the qualitative detection of SARS-CoV-2 Antigen, Flu A&B Antigen, Respiratory Syncytial Virus (RSV) antigen in Nasopharyngeal swab. For professional in vitro diagnostic use only.

#### [INTENDED USE]

The COVID-19+Flu A&B+RSV Antigen Combo Rapid Test Cassette (Nasopharyngeal swab) is a rapid chromatographic immunoassay for the qualitative detection of SARS-CoV-2, Influenza A&B, RSV Antigen in Nasopharyngeal swab. It is intended to aid in the rapid differential diagnosis of COVID-19, Influenza A and B, RSV infections.

#### (SUMMARY)

The novel coronaviruses belong to the β genus. COVID-19 is an acute respiratory infectious disease. People are generally susceptible. Currently, the patients infected by the novel coronavirus are the main source of infection; asymptomatic infected people can also be an infectious source. Based on the current epidemiological investigation, the incubation period is 1 to 14 days, mostly 3 to 7 days. The main manifestations include fever, fatigue, and dry cough. Nasal congestion, runny nose, sore throat, myalgia, and diarrhea are found in a few cases.

The COVID-19 Antigen Rapid Test is qualitatively detecting the presence of coronavirus COVID-19 antigen in Nasopharyngeal swab, providing results within 10 minutes. The test uses antibodies specific for coronavirus COVID-19 to selectively detect Nucleocapsid(N)protein of SARS-CoV-2 in Nasopharyngeal swab.

Influenza (commonly known as 'flu') is a highly contagious, acute viral infection of the respiratory tract. It is a communicable disease easily transmitted through the coughing and sneezing of aerosolized droplets containing live virus. Influenza outbreaks occur each year during the fall and winter months. Type A viruses are typically more prevalent than type B viruses and are associated with most serious influenza epidemics, while type B infections are usually milder.

The gold standard of laboratory diagnosis is 14-day cell culture with one of a variety of cell lines that can support the growth of influenza virus. Cell culture has limited clinical utility, as results are obtained too late in the clinical course for effective patient intervention. Reverse Transcriptase Polymerase Chain Reaction (RT-PCR) is a newer method that is generally more sensitive than culture with improved detection rates over culture of 2-23%. However, RT-PCR is expensive, complex and must be performed in specialized laboratories.

Human respiratory syncytial virus (HRSV) is a syncytial virus that causes respiratory tract infections. It is a major cause of lower respiratory tract infections and hospital visits during infancy and childhood. A prophylactic medication, palivizumab, can be employed to prevent HRSV in preterm (under 35 weeks gestation) infants, infants with certain congenital heart defects (CHD) or bronchopulmonary dysplasia (BPD), and infants with congenital malformations of the airway. Treatment is limited to supportive care (e.g., C-PAP), including oxygen therapy. In temperate climates there is an annual epidemic during the winter months. In tropical climates, infection is most common during the rainy season

The COVID-19+Flu A&B+RSV Antigen Combo Rapid Test Cassette (Nasopharyngeal swab) is a rapid test to qualitatively detect the presence of SARS-CoV-2, Influenza A&B, RSV Antigen in Nasopharyngeal swab.

# [PRINCIPLE]

The COVID-19+Flu A&B+RSV Antigen Combo Rapid Test Cassette (Nasopharyngeal swab) is a qualitative, lateral flow immunoassay for the detection of the RSV antigen, N protein of SARS-CoV-2, Influenza A and Influenza B nucleoproteins in Nasopharyngeal swab.

In this test, antibody specific to the Respiratory Syncytial Virus, antibody specific to the N protein of SARS-CoV-2, antibody specific to the Influenza A and Influenza B nucleoproteins are separately coated on the test line regions of the test cassette. During testing, the extracted specimen reacts with the antibody to N protein of antibody specific to the Respiratory Syncytial Virus, SARS-CoV-2, Influenza A and/or Influenza B nucleoproteins, that are coated onto particles. The mixture migrates up the membrane to react with the antibody specific to the Respiratory Syncytial Virus. antibody to N protein of SARS-CoV-2, Influenza A and/or Influenza B nucleoproteins on the membrane and generate one colored line in the test regions. The presence of this colored line of the test regions indicates a positive result. To serve as a procedural control, a colored line will always appear in the control region if the test has performed

# [REAGENTS]

The test cassette contains anti-Respiratory Syncytial Virus particles, anti-SARS-CoV-2 Nucleocapsid protein particles, anti-Influenza A&B virus particles and anti-Respiratory Syncytial Virus, anti-SARS-CoV-2 Nucleocapsid protein, anti- Influenza A & B Virus

coated on the membrane.

#### [PRECAUTIONS]

Please read all the information in this package insert before performing the test.

- 1. For professional in vitro diagnostic use only. Do not use after the expiration date.
- 2. The test should remain in the sealed pouch until ready to use.
- 3. All specimens should be considered potentially hazardous and handled in the same manner as an infection agent.
- 4. The used test should be discarded according to the local regulations.
- 5. Avoid using bloody samples.
- 6. Wear gloves when handling the samples, avoid touching the reagent membrane and sample well.

#### **[STORAGE AND STABILITY]**

through the expiration date printed on the sealed pouch. The test must remain in the sealed bouch until use. DO NOT FREEZE. Do not use beyond the expiration date.

#### **[SPECIMEN COLLECTION AND PREPARATION]**

#### Nasopharvngeal swab

Insert swab through the nostril parallel to the palate (not upwards) until resistance is encountered or the distance is equivalent to that from the ear to the nostril of the patient, indicating contact with the nasopharynx, swab should reach depth equal to distance from nostrils to outer opening of the ear. Gently rub and roll the swab. Leave swab in place for several seconds to absorb secretions. Slowly remove swab while rotating it. Specimens can be collected from both sides using the same swab, but it is not necessary to collect specimens from both sides if the swab is saturated with fluid from the first collection. If a deviated septum or blockage create difficulty in obtaining the specimen from one nostril, use the same swab to obtain the specimen from the

# Do not return the Nasopharyngeal swab to the original paper packaging.

• For best performance, direct Nasopharyngeal swabs should be tested as soon as possible after collection.



# [MATERIALS]

#### Materials provided Test cassettes **Extraction Buffer Tubes** Positive Control Swab

Negative Control Swab Workstation

Sterile Swabs Package Insert

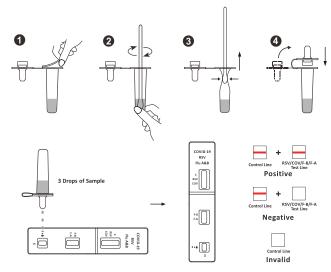
# Materials required but not provided

Timer

# [DIRECTIONS FOR USE]

Allow the test, specimen, extraction buffer to equilibrate to room temperature (15-30℃) prior to testing.

- 1. Remove the test cassette from the sealed foil pouch and use it within one hour. Best results will be obtained if the assay is performed immediately after opening the foil pouch.
- 2. Tear the aluminum foil on the extraction buffer tube. See illustration 1.
- 3. Place the swab specimen in the extraction buffer tube. Rotate the swab for approximately 10 seconds while pressing the head against the inside of the extraction buffer tube body to release the antigen in the swab. See illustration 2.
- 4. Remove the swab while squeezing the swab head against the inside of the individual tube as you remove it to expel as much liquid as possible from the swab. See illustration 3
- 5. Fit the dropper tip on top of the extraction buffer tube. Place the test cassette on a clean and flat surface. Do not move the test cassette during the test. See illustration
- 6. Hold the tube vertically and transfer 3 drops of the sample solution (approx.150µL) to the sample well and then start the timer. Read the result at 10 minutes. Do not interpret the result after 20 minutes.
- 7. Please dispose off the swab, extraction buffer tube and test cassette in the disposal bag provided inside the test kit package. Wash your hand after the test.



#### [INTERPRETATION OF RESULTS]

(Please refer to the illustration above)

POSITIVE: \* A colored line appears in the Control region (C) and colored line(s) appears in the Test region (T (RSV, COV and F-B.F-A). A positive result in the RSV T region indicates that RSV was detected in the specimen, a positive result in the COV T region indicates that SARS-CoV-2 was detected in the specimen, a positive result in the F-B region indicates that Influenza B antigen was detected in the specimen, a positive result in the F-A region indicates that Influenza A antigen was detected in the

\*NOTE: The shade of the colored line(s) in the Test region(T (RSV, COV and F-B,F-A) may vary. The result should be considered positive whenever there is even a faint line. **NEGATIVE:** One colored line appears in the control line region (C). No line appears in the test line region (T (RSV, COV and F-B,F-A). A negative result indicates that RSV. COVID-19, Influenza A&B are not present in the specimen, or is present below the detectable level of the test

INVALID: Control line fails to appear, Insufficient specimen volume or incorrect procedural techniques are the most likely reasons for control line failure. Review the procedure and repeat the test with a new test. If the problem persists, discontinue using the test kit immediately and contact your local distributor.

# [QUALITY CONTROL]

# INTERNAL QUALITY CONTROL:

A procedural control is included in the test. A colored line appearing in the control line region (C) is considered an internal procedural control. It confirms adequate membrane wicking.

**EXTERNAL QUALITY CONTROL:** It is recommended that positive external controls be run every kit, and as deemed necessary by your internal laboratory procedures. External positive controls are supplied in the kit. Alternatively, other RSV, SARS-CoV-2. FLU A&B reference strains may be used as external controls. Some commercial controls may contain interfering preservatives; therefore, other commercial controls are not recommended

# PROCEDURE FOR EXTERNAL QUALITY CONTROL TESTING:

- 1. Tear the aluminum foil on the extraction buffer tube. See illustration 1.
- 2. Place the swab specimen in the extraction buffer tube. Rotate the swab for approximately 10 seconds while pressing the head against the inside of the extraction buffer tube body to release the antigen in the swab. See illustration 2.
- 3. Remove the swab while squeezing the swab head against the inside of the individual tube as you remove it to expel as much liquid as possible from the swab. See illustration 3.
- 4. Fit the dropper tip on top of the extraction buffer tube. Place the test cassette on a clean and flat surface. Do not move the test cassette during the test. See illustration
- 5. Hold the tube vertically and transfer 3 drops of the sample solution (approx.150µL) to the sample well and then start the timer. Read the result at 10 minutes. Do not interpret the result after 20 minutes.
- 6. Please dispose off the swab, extraction buffer tube and test cassette in the disposal bag provided inside the test kit package. Wash your hand after the test.

# [LIMITATIONS]

1. The COVID-19+Flu A&B+RSV Antigen Combo Rapid Test Cassette

(Nasopharyngeal swab) is for professional in vitro diagnostic use only. The test should be used for the detection of RSV, SARS-CoV-2, Influenza A&B Antigen Nasopharyngeal swab. Neither the quantitative value nor the rate of increase in SARS-CoV-2 virus, Influenza A&B virus, Respiratory Syncytial Virus concentration can be determined by this qualitative test.

- The accuracy of the test depends on the quality of the swab sample. False negatives may result from improper sample collection or storage.
- The COVID-19+Flu A&B+RSV Antigen Combo Rapid Test Cassette (Nasopharyngeal swab) will only indicate the presence of SARS-CoV-2, Influenza A&B, RSV in the specimen from both viable and non-viable SARS-CoV-2 coronavirus, Influenza A and B strains Respiratory Syncytial Virus.
- As with all diagnostic tests, all results must be interpreted together with other clinical information available to the physician.
- A negative result obtained from this kit should be confirmed by PCR. A negative result may be obtained if the concentration of the SARS-CoV-2, Influenza A&B virus, Respiratory Syncytial Virus present in the swab is not adequate or is below the detectable level of the test.
- Excess blood or mucus on the swab specimen may interfere with test performance and may yield a false positive result.
- Positive result for COVID-19, Influenza A, Influenza B, Respiratory Syncytial Virus
  do not preclude an underlying co-infection with another pathogen, therefore the
  possibility of an underlying bacterial infection should be considered.
- Negative results do not rule out SARS-CoV-2, Influenza A, Influenza B infection, RSV particularly in those who have been in contact with the virus. Follow-up testing with a molecular diagnostic should be considered to rule out infection in these individuals.
- The use of over-the-counter and prescription nasal sprays at high concentrations can interfere with results, leading to either invalid or incorrect test results.
- COVID-19 positive results may be due to current infection with acute non-SARS-CoV-2 coronavirus strains, such as coronavirus HKU1, NL63, OC43 or 299F
- Results from antigen testing should not be used as the sole basis to diagnose or exclude SARS-CoV-2, Influenza A &B, RSV infection or to inform infection status.
- 12. Extraction reagent has the ability to kill the virus, but it cannot inactivate 100% of the virus. The method of inactivating the virus can be referred to: what method is recommended by WHO/CDC, or it can be handled according to local regulations.

#### [PERFORMANCE CHARACTERISTICS]

### Sensitivity, Specificity and Accuracy

The COVID-19+Flu A&B+RSV Antigen Combo Rapid Test Cassette (Nasopharyngeal swab) has been evaluated with specimens obtained from the patients. PCR is used as the reference method for the COVID-19+Flu A&B+RSV Antigen Combo Rapid Test Cassette (Nasopharyngeal swab). Specimens were considered positive if PCR indicated a positive result.

indicated a positive result.							
COVID-19+Flu A&B+RSV		RSV		COVID-19			
Antigen Combo Rapid Test	PC	R	Total	PCR		Total	
Cassette	Positive	ive Negative		Positive	Negative	Total	
Positive	71	3	74	80	1	81	
Negative	4	448	452	3	442	445	
Total	75	451	526	83	443	526	
Relative Sensitivity	94.7%(95%	CI*:86.9%~9	98.5%)	96.4%(95%CI*:89.8%~99.2%			
Relative Specificity	99.3%(95%CI*:98.1%~99.9%)			99.8%(95%CI*:98.7%~100.0%)			
Accuracy	98.7%(95%CI*:97.3%~99.5%)			99.2%(95%CI*:98.1%~99.8%)			
COVID-19+Flu A&B+ RSV	FLU B			FLUA			
Antigen Combo Rapid Test	PCR To			PCR		Total	
Cassette	Positive	Negative	Total	Positive	Negative	Total	
Positive	53	8	61	101	5	106	
Negative	3	462	465	6	414	420	
Total	56	470	526	107	419	526	
Relative Sensitivity	94.6%(95%CI*:85.1%~98.9%)			94.4%(95%CI*:88.2%~97.9%)			
Relative Specificity	98.3%(95%CI*:96.7%~99.3%) 98.8%(95%CI*:97.2%~			~99.6%)			
Accuracy	97.9%(95%CI*:96.3%~99.0%) 97.9%(95%CI*:96.3%~99.0%			~99.0%)			
	Dotooti	an Limit of	DCV				

#### **Detection Limit of RSV**

The minimum detection limit of RSV Antigen Rapid Test is as follows:

Subtype	VP/mL
A2	1.12X10 <sup>2</sup> TCID <sub>50</sub> /mL
B WV/14617/85	3.67 X 10⁴ PFU/mL
18537	32 PFU/mL

#### Detection Limit of COVID-19

The LOD for the COVID-19 Antigen Rapid Test (Nasopharyngeal swab) was established using limiting dilutions of a viral sample inactivated. The material (ZeptoMetrix, 0810587CFHI) was supplied at a concentration of  $1.15 \times 10^7 TCID_{50}/mL$ . The Estimated LOD is  $1000 TCID_{50}/mL$ .

 0 200	Strain	
Strain		Concentration of LOD

A/Mal/302/54	5×10 <sup>4</sup> CEID <sub>50</sub> /mL
A/NWS/33	7.4×10 <sup>4</sup> CEID <sub>50</sub> /mL
A/Hong Kong/8/68	1.6×10 <sup>4</sup> CEID <sub>50</sub> /mL
A/Aichi/2/68	2.3×10 <sup>4</sup> CEID <sub>50</sub> /mL
A/Port Chalmers/1/73	2.5×105CEID50/mL
A/New Jersey/8/76	8.9×10 <sup>4</sup> CEID <sub>50</sub> /mL
A/WS/33	1.6×10 <sup>4</sup> CEID <sub>50</sub> /mL
A/Anhui/1/2013	95ng HA/mL
Hong Kong/2671/2019	64ng HA/mL
Guangdong-Maonan/SWL1536/2019	770ng HA/mL
Influenza Antigen A/mallard/Netherlands/12/2000 (H7N3)	1.55µg HA/mL
Influenza Antigen A/Cambodia/RO405050/2007 (H5N1)	930ng HA/mL
B/Lee/40	1.6×10 <sup>4</sup> CEID <sub>50</sub> /mL
B/Brigit	1.6×105CEID50/mL
B/R5	1.6×10 <sup>3</sup> CEID <sub>50</sub> /mL
R/Russia/69	8.9×10 <sup>3</sup> CEID <sub>50</sub> /mL
B/Hong Kong/5/72	8.9×10 <sup>4</sup> CEID <sub>50</sub> /mL
Influenza B Virus B/Wisconsin/1/2010	2.8×10 <sup>3</sup> CEID <sub>50</sub> /mL
Influenza B Virus B/Florida/78/2015	7.8×10 <sup>3</sup> TCID <sub>50</sub> /mL
Phuket/3073/2013	0.6µg HA/mL
Washington/02/2019	0.76µg/mL

#### **Cross Reactivity**

The COVID-19 Antigen Rapid Test (Nasopharyngeal swab) has been tested for Influenza A virus, Influenza B virus, Adenovirus, Coxsackie virus, Parainfluenza Virus Type1, Parainfluenza Virus Type2, Parainfluenza Virus Type4a, Enterovirus, Mumps virus, Respiratory syncytial virus, Rhinovirus, Bordetella pertussis, Haemophilus parainfluenzae, Staphylococcus aureus, Streptococcus agalactiae, Neisseria meningitides, Streptococcus sp. group A, Streptococcus sp. group B, Streptococcus sp. group C, Candida albicans, Human Metapneumovirus (hMPV), Legionella pneumophila, Mycobacterium tuberculosis, Mycoplasma pneumoniae, Pneumocystis jirovecii(PJP)-S cerevisiae Recombinant, Pseudomonas aeruginosa, Staphylococcus epidermis, Streptococcus pneumoniae, Streptococcus pyogenes, Streptococcus salivarius, Human coronavirus 229E, Human coronavirus OC43, Human coronavirus NL63, MERS-coronavirus positive specimens. The results showed no cross reactivity.

The FLU A&B Antigen Rapid Test (Nasopharyngeal swab) has been tested for Adenovirus, Coxsackie virus, Cytomegalovirus, Parainfluenza Virus Type1,2,3,4a, Enterovirus, Mumps virus, Respiratory syncytial virus, Rhinovirus, Bordetella pertussis, Haemophilusparainfluenzae, Staphylococcus aureus, Streptococcus agalactiae, Neisseria meningitides, Streptococcus sp. group A, B, C. The results showed no cross reactivity.

The RSV Antigen Rapid Test (Nasopharyngeal swab) has been tested for Adenovirus, Coxsackie virus, Cytomegalovirus, Parainfluenza Virus Type1,2,3,4a, Enterovirus, Mumps virus, Influenza A virus, Influenza B virus, Rhinovirus, Bordetella pertussis, Haemophilusparainfluenzae, Staphylococcus aureus, Streptococcus agalactiae, Neisseria meningitides, Streptococcus sp. group A, B, C. The results showed no cross reactivity.

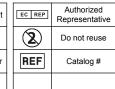
### [BIBLIOGRAPHY]

- Weiss SR, Leibowitz JL. Coronavirus pathogenesis. Adv Virus Res 2011; 81:85-164
- Cui J, Li F, Shi ZL. Origin and evolution of pathogenic coronaviruses. Nat Rev Microbiol 2019: 17:181-192.
- Su S, Wong G, Shi W, et al. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. TrendsMicrobiol 2016; 24:490-502.
- Williams, KM, Jackson MA, Hamilton M. (2002) Rapid Diagnostic Testing for URIs in Children; Impact on Physician Decision Making and Cost. Infec. Med. 19(3): 1102-111
- Betts, R.F. 1995. Influenza virus, p. 1546-1567. In G.L. Mandell, R.G. Douglas, Jr. and J.E. Bennett (ed.), Principle and practice of infectious diseases, 4th ed. Churchill Livingstone, Inc., New York, N.Y.
- WHO recommendations on the use of rapid testing for influenza diagnosis, World Health Organisation, July 2005.
- Norihiko KUBO, Hideyuki IKEMATSU, Shigeki NABESHIMA: Evaluation of an Immunochromatography TestKit for Rapid Diagnosis of Influenz, Kansenshogaku Zasshi, 2003,77:1007~1014.
- Michimaru HARA, Shinichi TAKAO, Shinji FUKUDA, Yukie SHIMAZU, Masaru KUWAYAMA and Kazuo MIYAZAKI: Comparison of Four Rapid Diagnostic Kits Using Immunochromatography to Detect Influenza B Viruses, Kansenshogaku Zasshi. 2005.79:803~811.
- Rezaee, F., Linfield, D.T., Harford, T. J., & Piedimonte, G. (2017). Ongoing developments in RSV prophylaxis: a clinician's analysis. Current Opinion in Virology, 24, 70-78.
- 10. F M Moesker, J J A van Kampen, G Aron, M Schutten, D A M C van de Vijver, M P G Koopmans, A D M E Osterhaus, P L A Fraaij. Diagnostic performance of influenza viruses and RSV rapid antigen detection tests in children in tertiary care.

- Journal of Clinical Virology, 2016, 79, 12-17.
- Yen, A. B., Demmler-Harrison, G. J. Rapid Antigen Testing to Detect Respiratory Syncytial Virus Performs Well in Neonates. The Pediatric Infectious Disease Journal, 2011, 30(3), 234-237.
- Slinger, R., Milk, R., Gaboury, I., & Diaz-Mitoma, F. Evaluation of the QuickLab RSV Test, a New Rapid Lateral-Flow Immunoassay for Detection of Respiratory Syncytial Virus Antigen. Journal of Clinical Microbiology, 2004, 42(8), 3731-3733.
- 13. Urs F Greber, Justin W Flatt. Adenovirus Entry: From Infection to Immunity. Annu Rev Virol. 2019 Sep 29;6(1):177-197.
- José Luiz Proenca-Modena, Ricardo de Souza Cardoso, Miriã Ferreira Criado, etc. Human adenovirus replication and persistence in hypertrophic adenoids and palatine tonsils in children. J Med Virol. 2019 Jul;91(7):1250-1262.

# Index of Symbols

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[]i	Consult Instruction for use	Σ	Tests per kit	EC REP
IVD	For in vitro diagnostic use only		Use by	<b>(2)</b>
2°C - 30°C	Store between 2-30°C	LOT	Lot Number	REF
<b>®</b>	Do not use if package is damaged			







EC REP

Shanghai International
Holding Corp. GmbH (Europe)
Eiffestrasse 80,
20537 Hamburg, Germany